



# Metabolic Engineering

Plant Metabolic Engineering  
under Construction

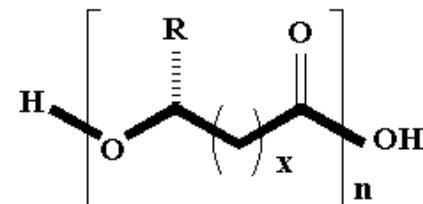
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Iowa State University



- Food for Humans
- Feed for Animals
- Fiber



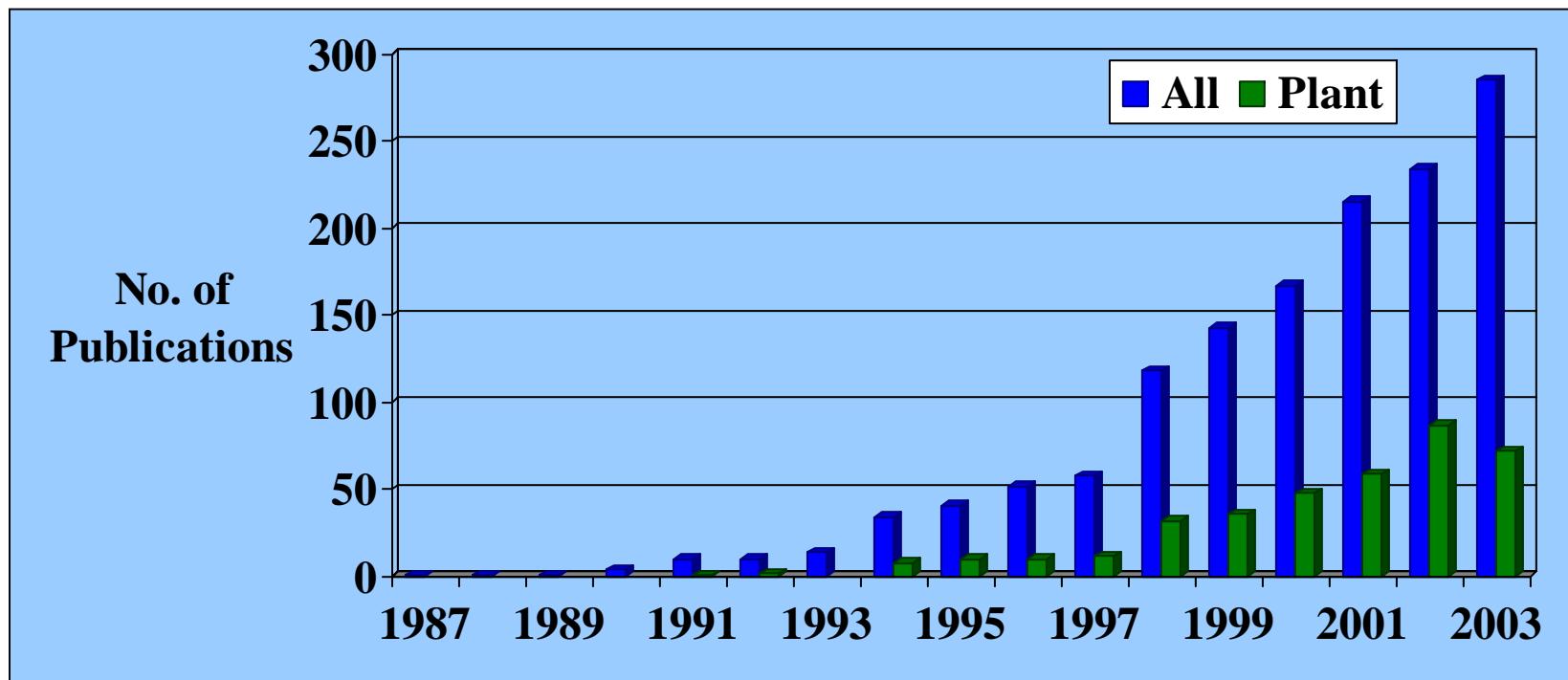
- Fuel
- Pharmaceuticals
- Feedstocks for the Chemical Industry



7th F - Phytoremediation



# Publications\* containing “Metabolic Engineering”



\*SciFinder Scholar

↑  
*Science*  
articles

↑  
*Metabolic Engineering*  
journal



1999

Journal  
dedicated to  
Metabolic  
Engineering  
is founded

JAN 2002

ME issue  
dedicated to  
Plant Metabolic  
Engineering

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# METABOLIC ENGINEERING

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**A Futile Cycle of Production and Oxidation of Lauric Acid in Transgenic Canola**

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# Synthesis Research Needs

- Necessary genes cloned
- Transform plants with multiple genes
- Organ, cell-specific, or inducible promoters



# Analysis Research Needs

- **Measurement tools**
  - protein and metabolite levels
  - metabolic fluxes
  - simultaneously and as quickly as possible
- **Models to analyze networks**
- **Uncertainty in measurements tied to predictions/estimates**





# **Overproduction of Tryptophan and Indole Alkaloids in *C. roseus***

**Project Number (BES 9906978, 0224600)**

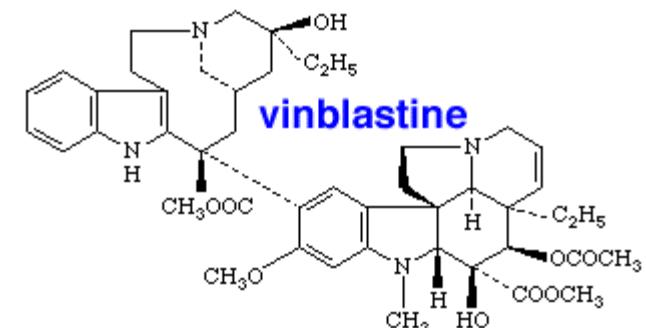
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Christy Peebles (Rice), Guy Sander (ISU)  
Dennis Hong (Rice & Univ. Minn)**



# *Catharanthus roseus*

- Therapeutic alkaloids
  - Vincristine & Vinblastine  $10^6$  \$/kg
    - anticancer
  - Ajmalicine & Serpentine
    - anti-hypertension
- Alkaloids 1% (DW)
- Chemical or microbial synthesis unfeasible



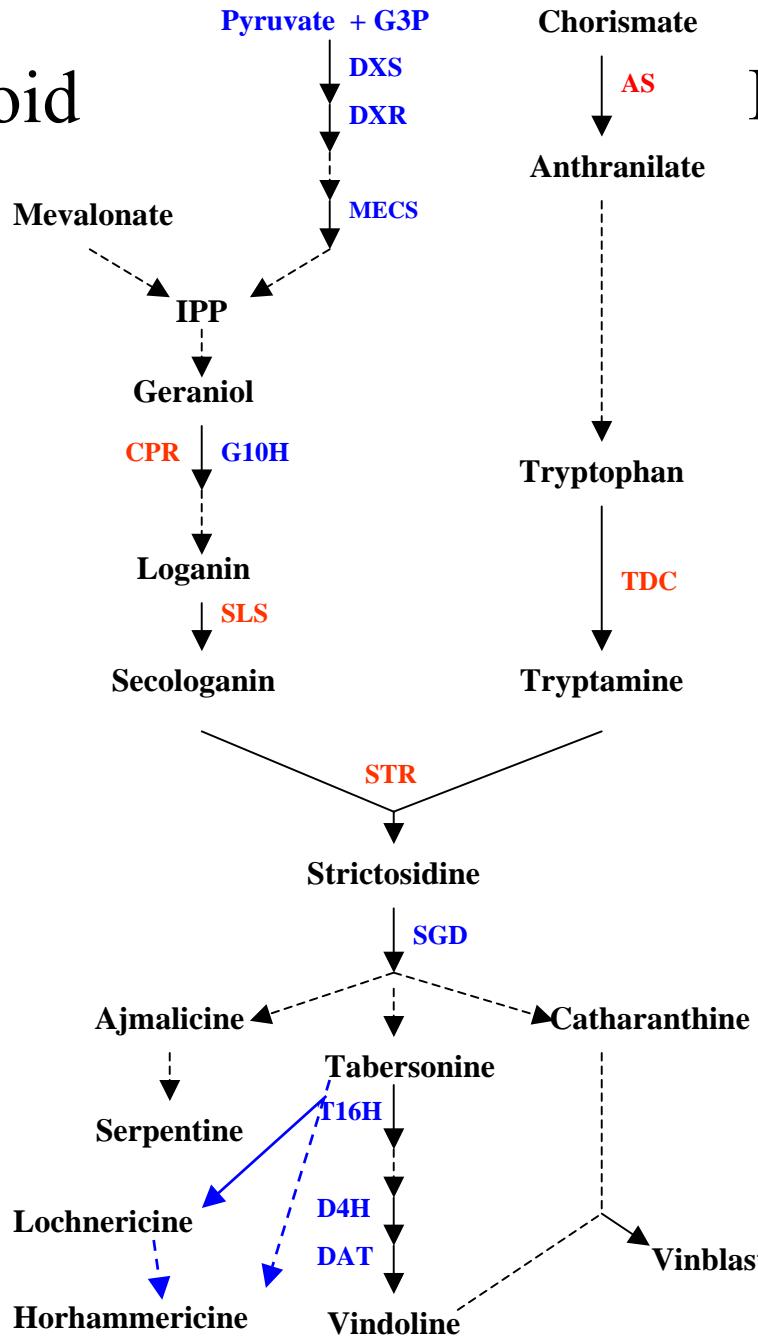


# Indole Alkaloids Pathways

**~1997**

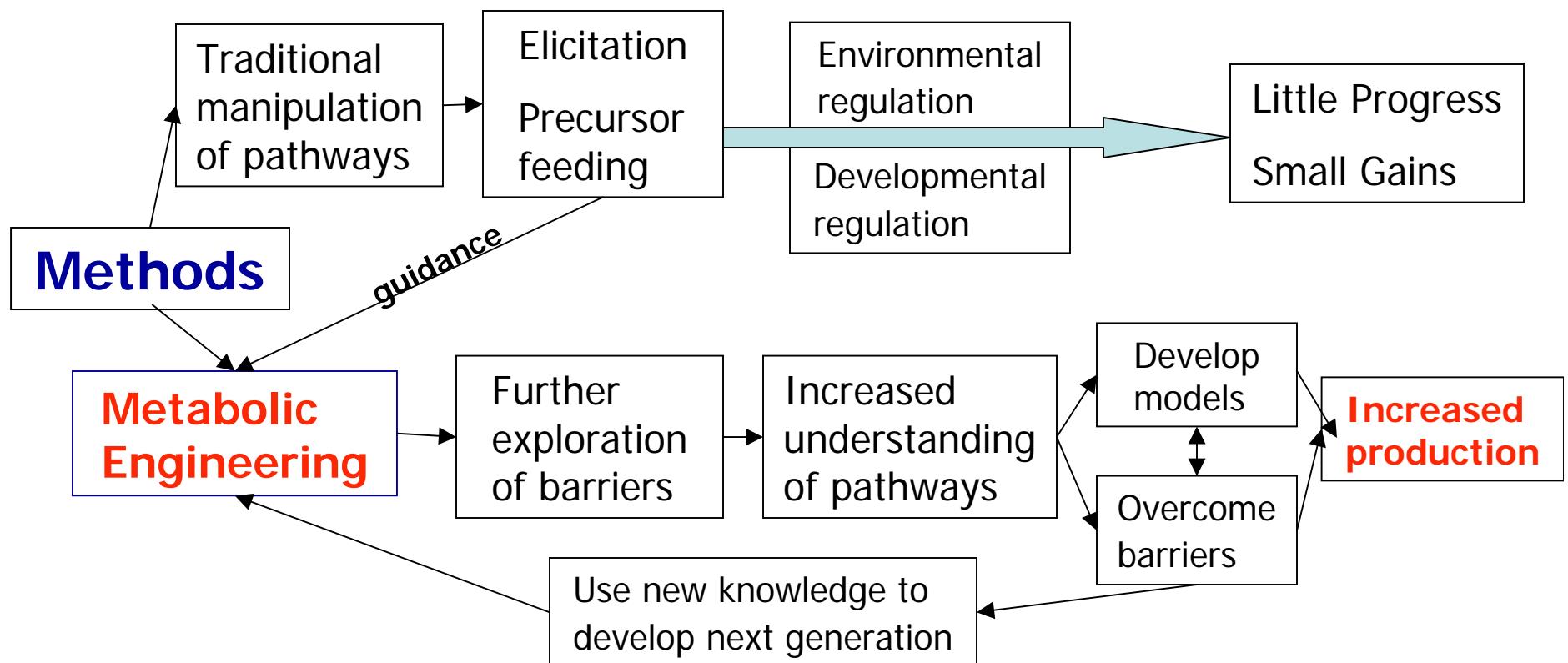
**2000**

## Terpenoid

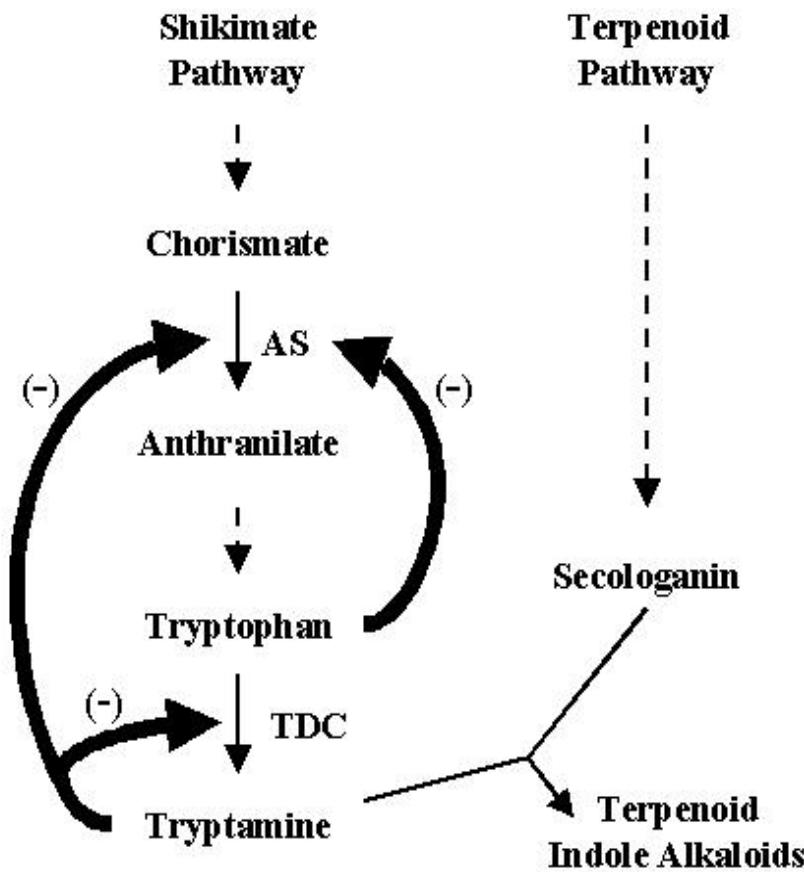


## Indole

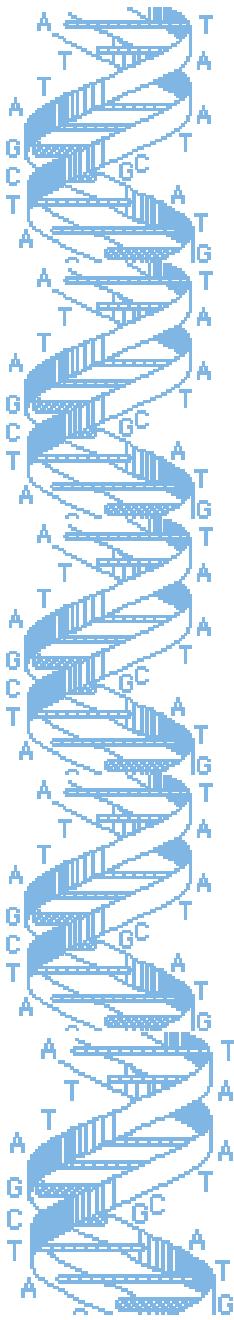
# Motivation



# Pathway Regulation by Anthranilate Synthase (AS)



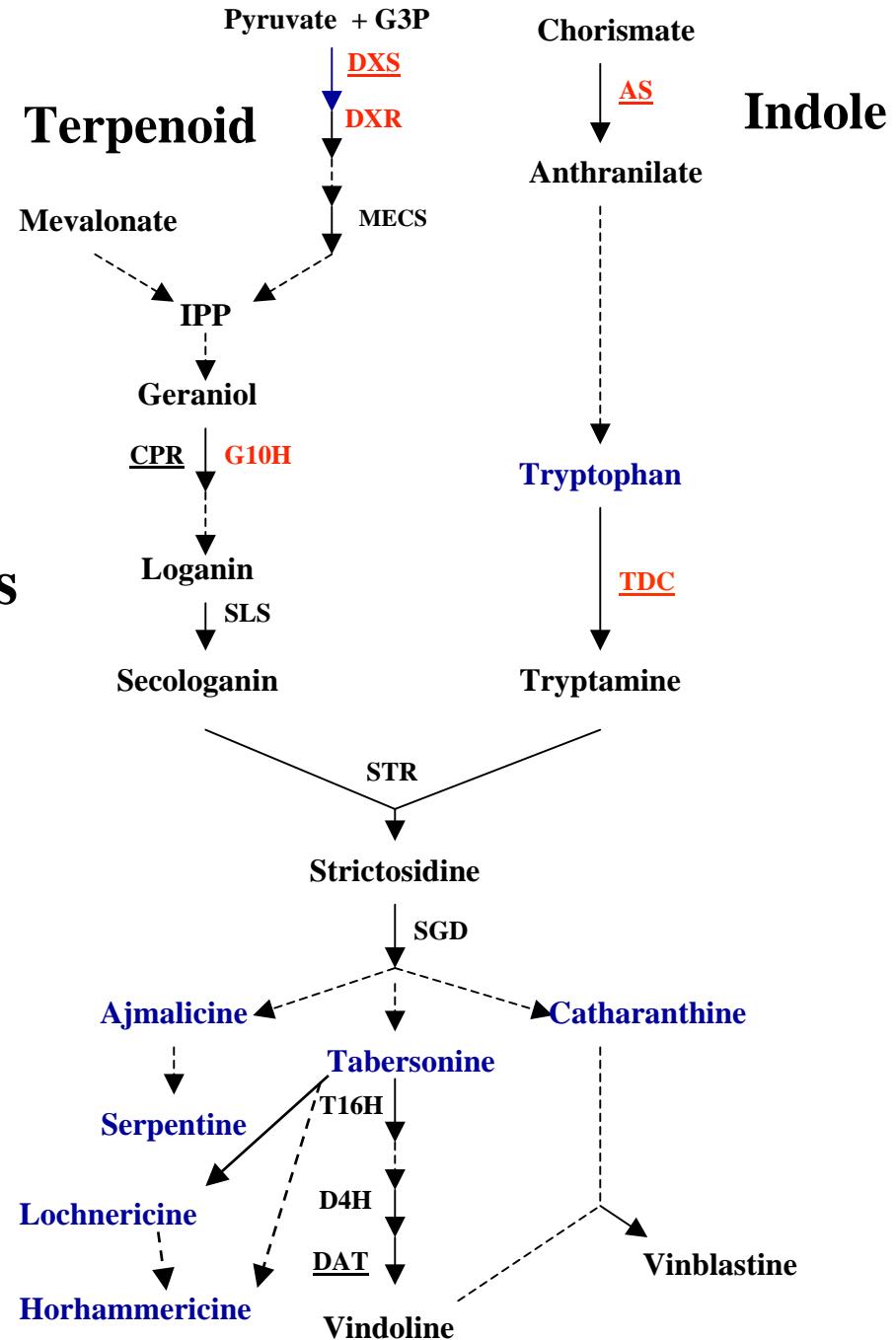
- **AS Activity**
  - $\alpha$  Feedback Inhibited by Tryptophan
  - $\beta$  unit?
- **Tryptophan Feeding**
  - 1.5x alkaloid levels (exponential growth)
- **Genetic Manipulations**
  - Feedback resistant *Arabidopsis AS $\alpha$* 
    - 3x tryptophan levels
  - TDC from *C. roseus*
    - Elevated tryptamine in cell culture



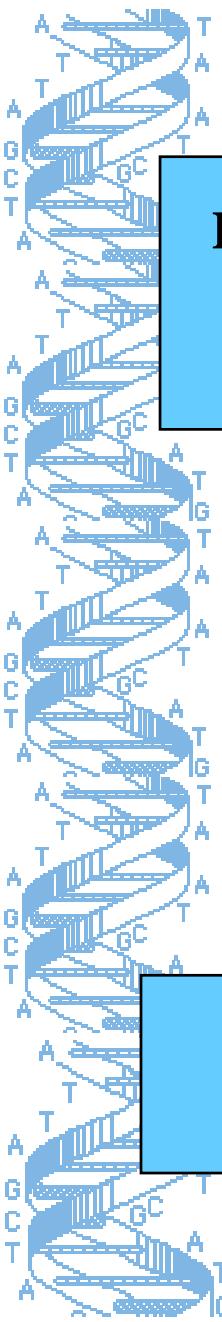
**Technical Objective**

- Engineer *C. roseus* hairy roots
- Overproduce tryptophan  
indole alkaloids

**Genetic targets: RED**  
**ORCA3 induced**



# Technical Approach



Develop an inducible promoter system in *C. roseus* hairy root cultures

NMR Flux Maps

Construct and characterize transgenic hairy root lines

Develop co-transformation techniques for insertion of multiple genes

Metabolic characterization of 1<sup>st</sup> generation lines

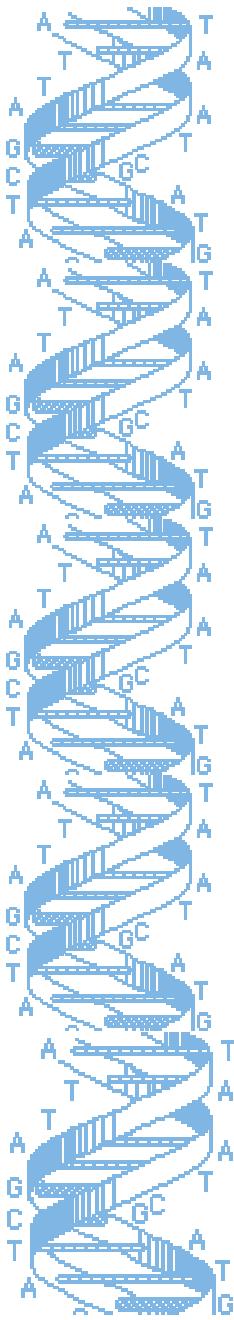
Construct and characterize 2<sup>nd</sup> generation lines



# Hairy root cultures

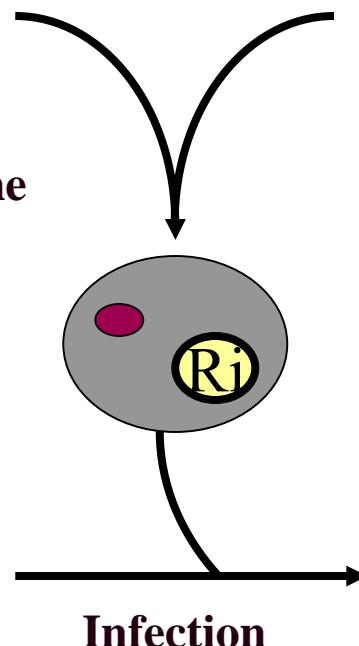
- Model system
- Transformed by Agrobacterium
- Increased genetic stability
- Fast & differentiated growth
- Higher alkaloid productivity





# Clone Generation

**Plasmid Construction  
in *E. coli***  
● Transgene

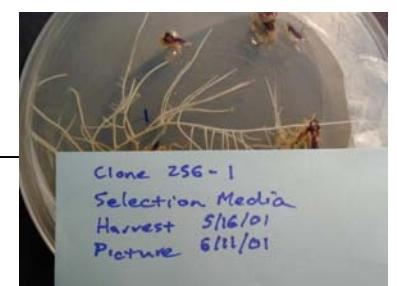


**ATCC 15834  
*A. rhizogenes***



**(6 weeks)**

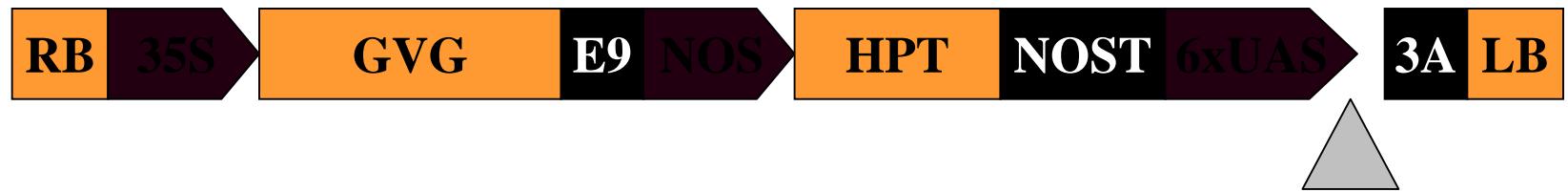
**Adapt to  
Liquid Media  
(12 weeks)**



**Selection Media  
(6 weeks)**



# Inducible-Promoter System

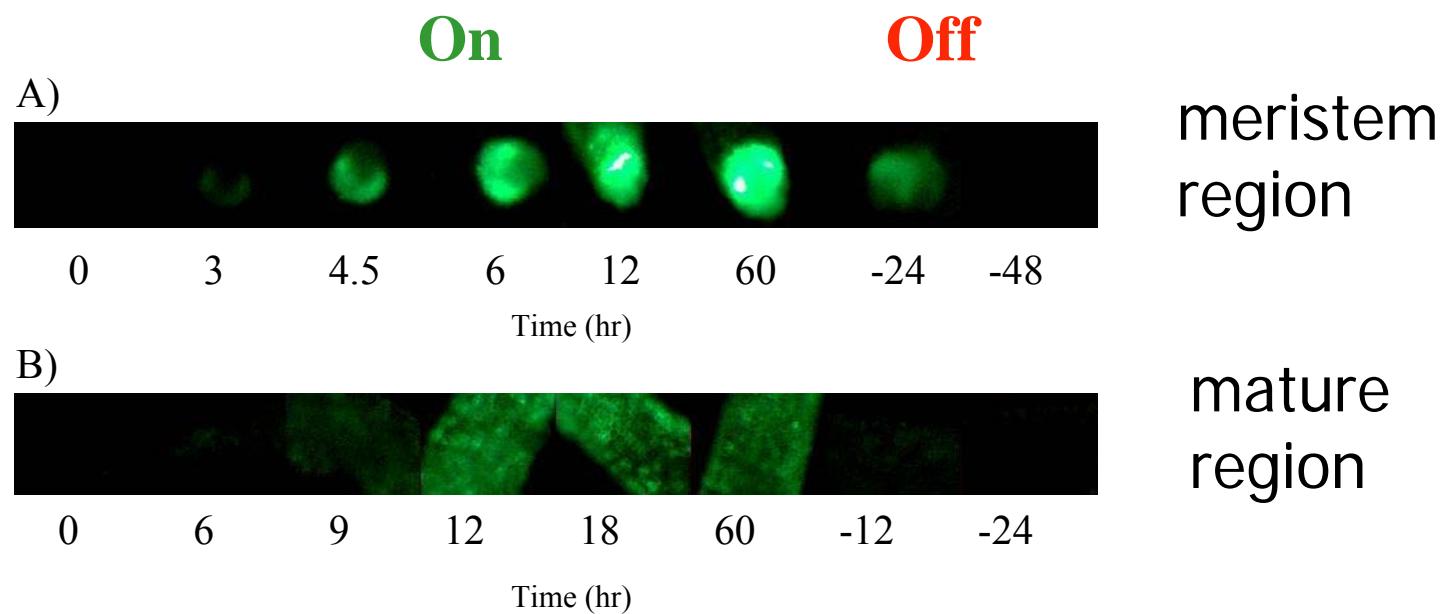


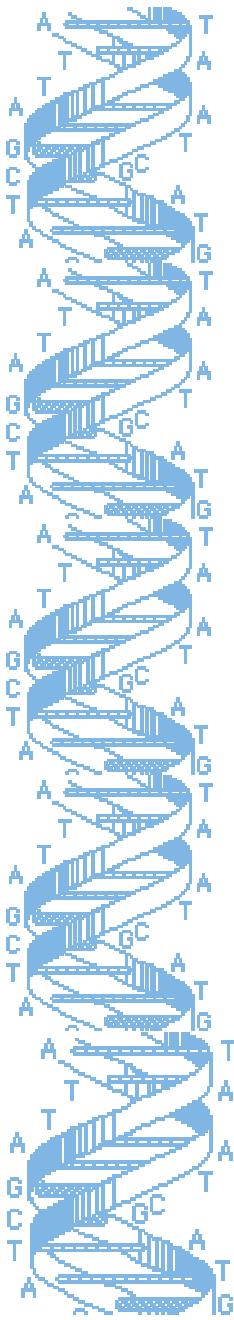
## Vertebrate Steroid Hormone Receptor System

- GVG Element  
(Glucocorticoid regulated transcription factor)
- Inducer - Dexamethasone (DEX)  
an artificial glucocorticoid hormone

# Inducible-Promoter System

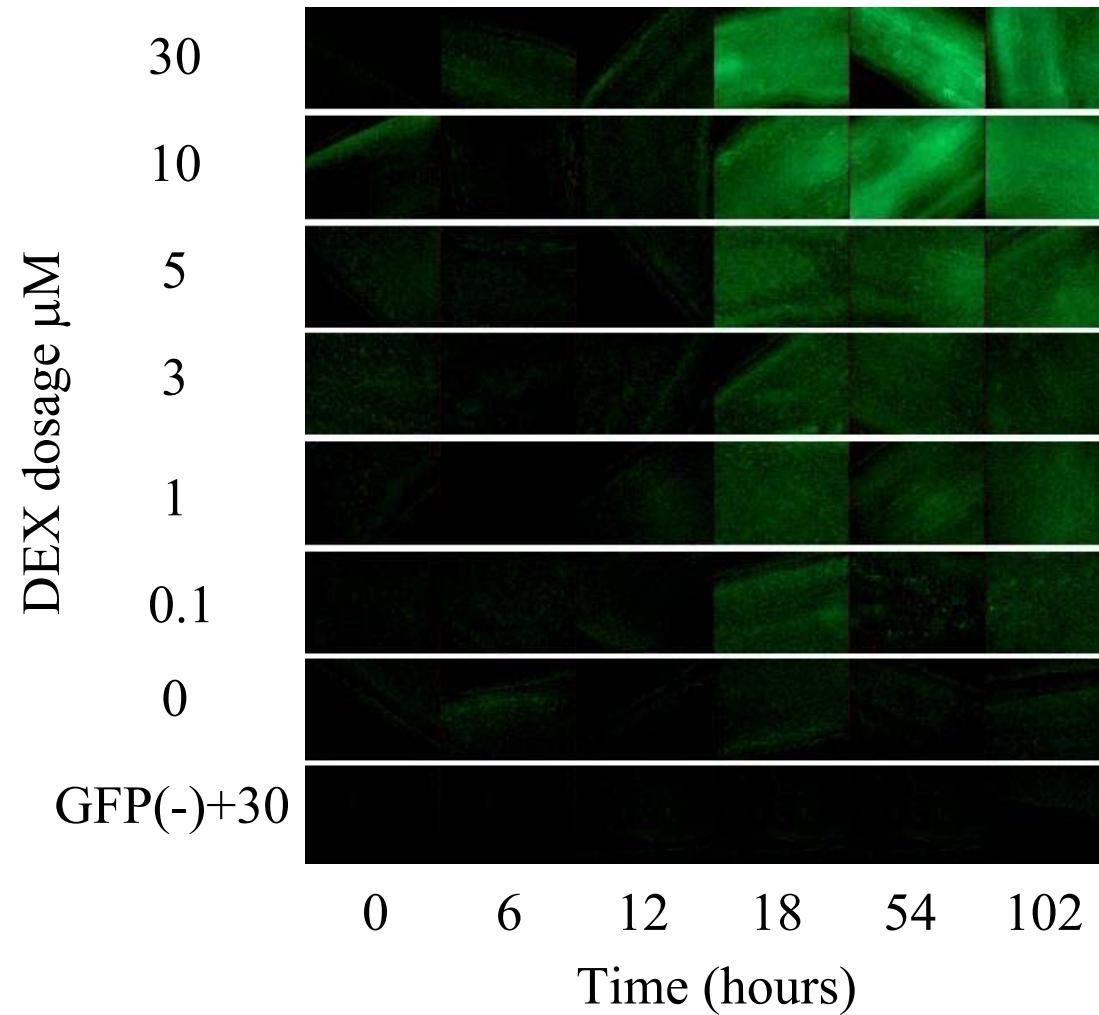
- Dexamethasone-inducible expression of green fluorescent protein (GFP)

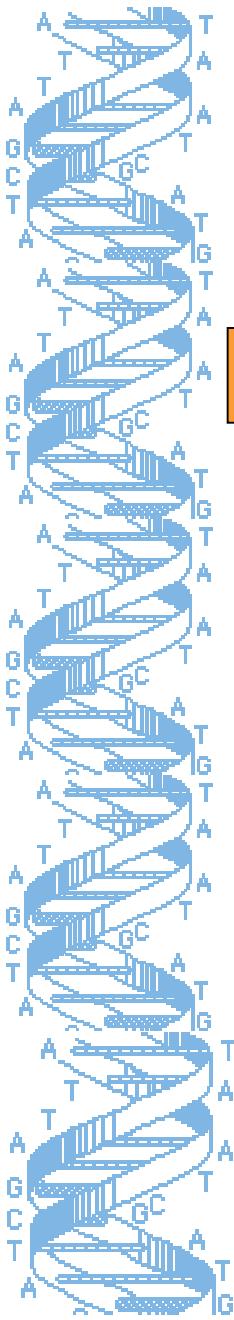




# Inducible-Promoter System

Dosage dependent response

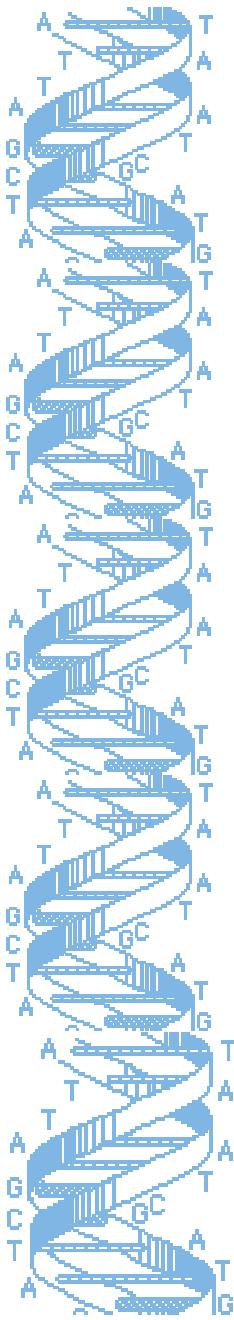




# AS $\alpha$ and TDC Clones

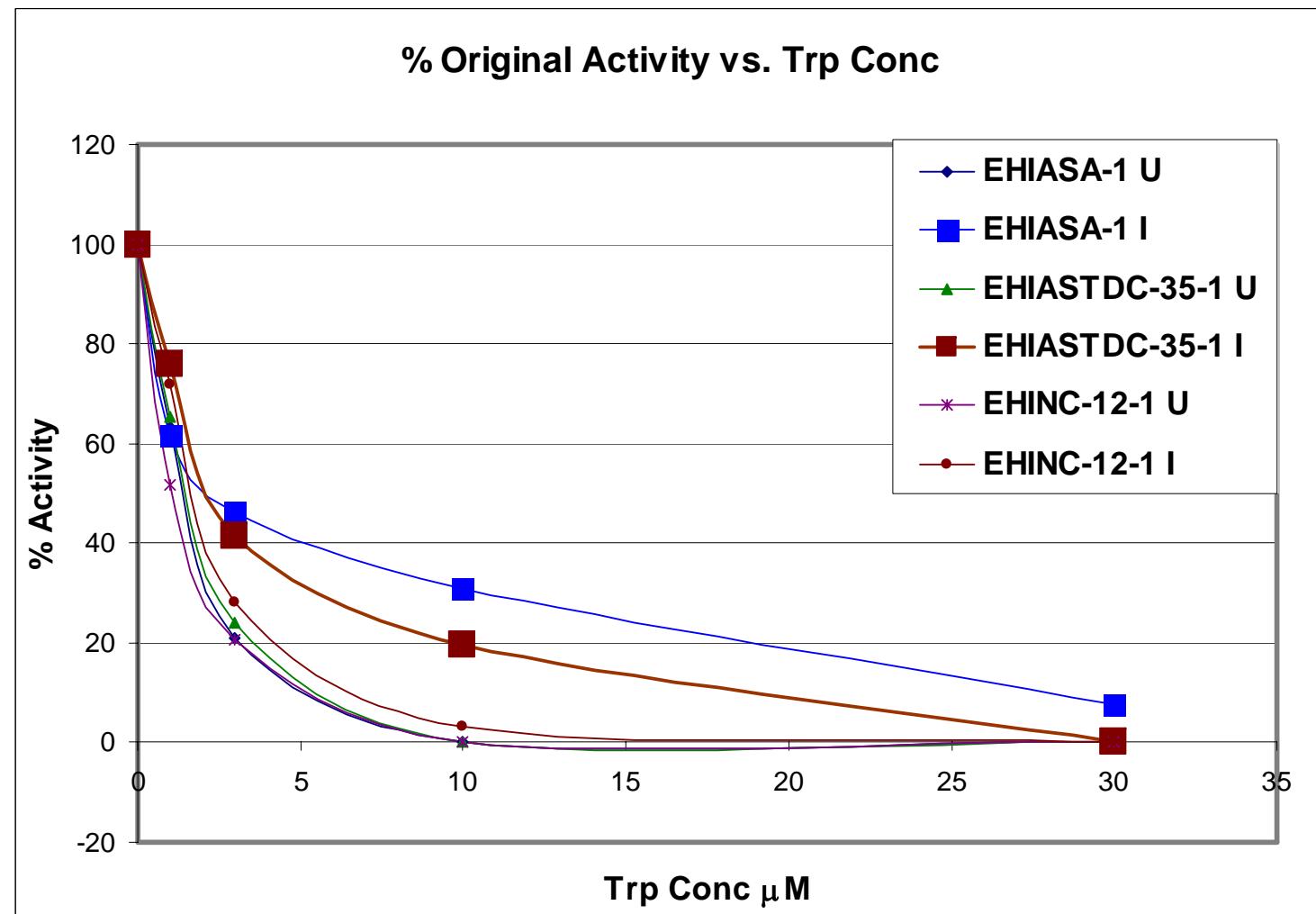


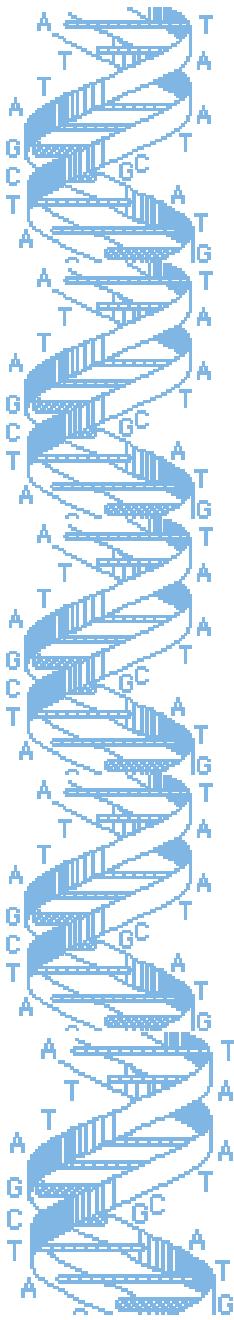
	Constitutive GVG	Inducible ASalpha	Inducible TDC
EHI <b>NC</b> -12-1	✓		
EHI <b>ASA</b> -1	✓	✓	
EHI <b>TDC</b> -15-2	✓		✓
EHI <b>ASTDC</b> -35-1	✓	✓	✓



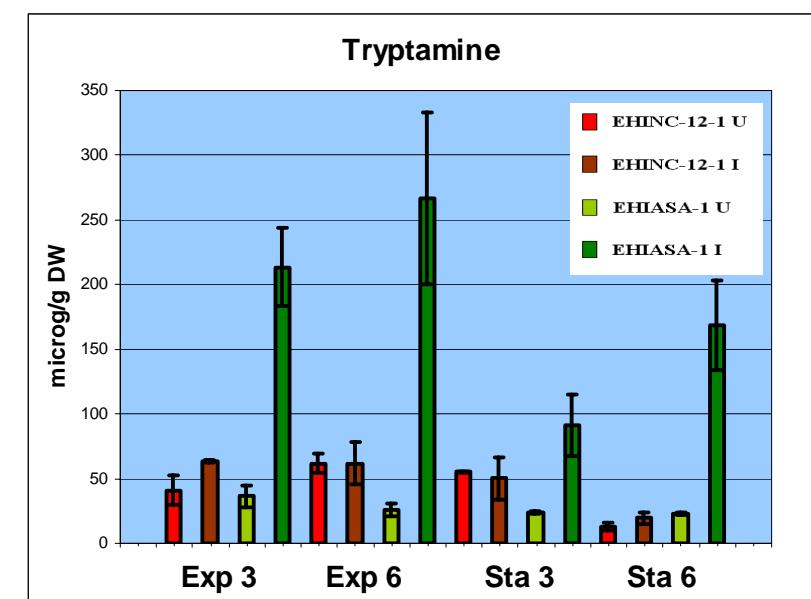
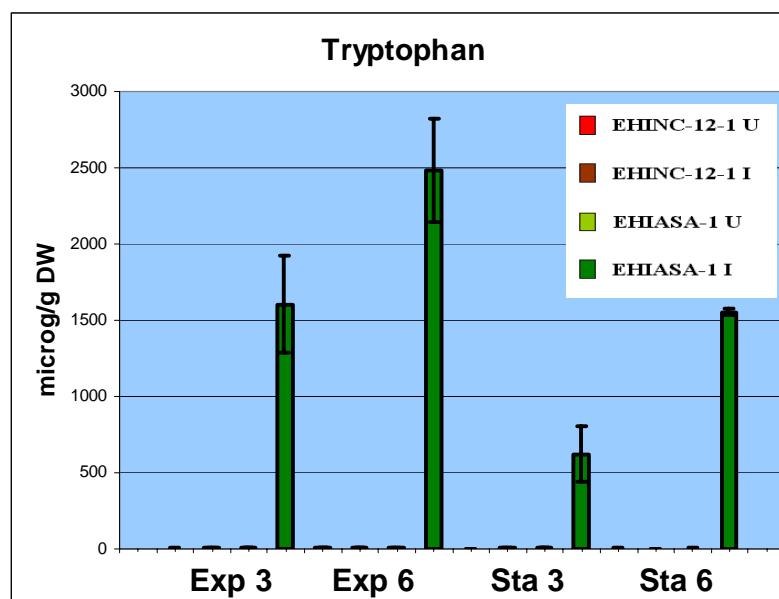
# Feedback Inhibited AS

Induced 3  $\mu\text{M}$  for 72 hours



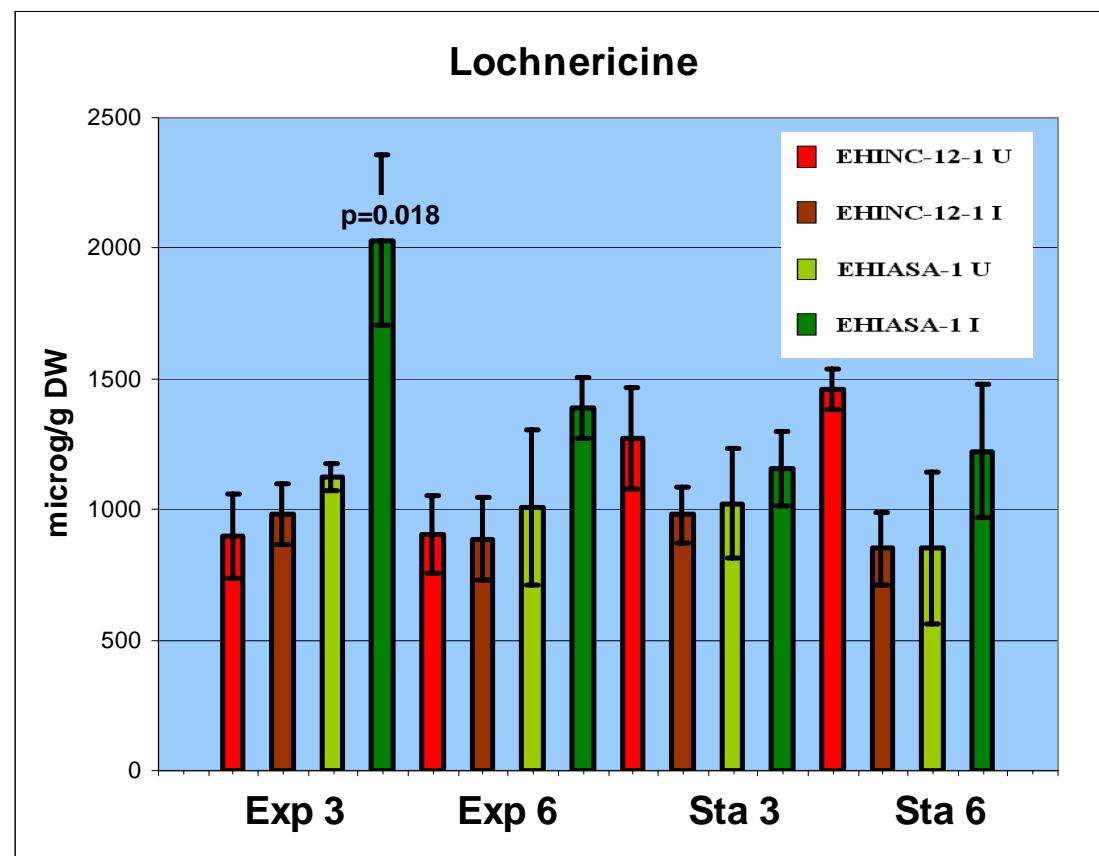


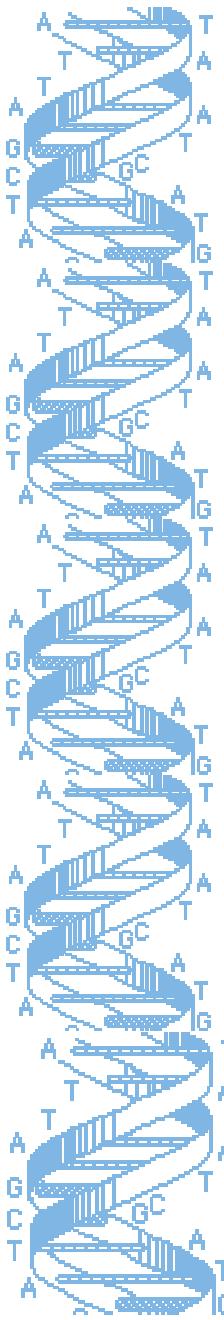
# AS $\alpha$ line – huge increases in tryptophan and tryptamine





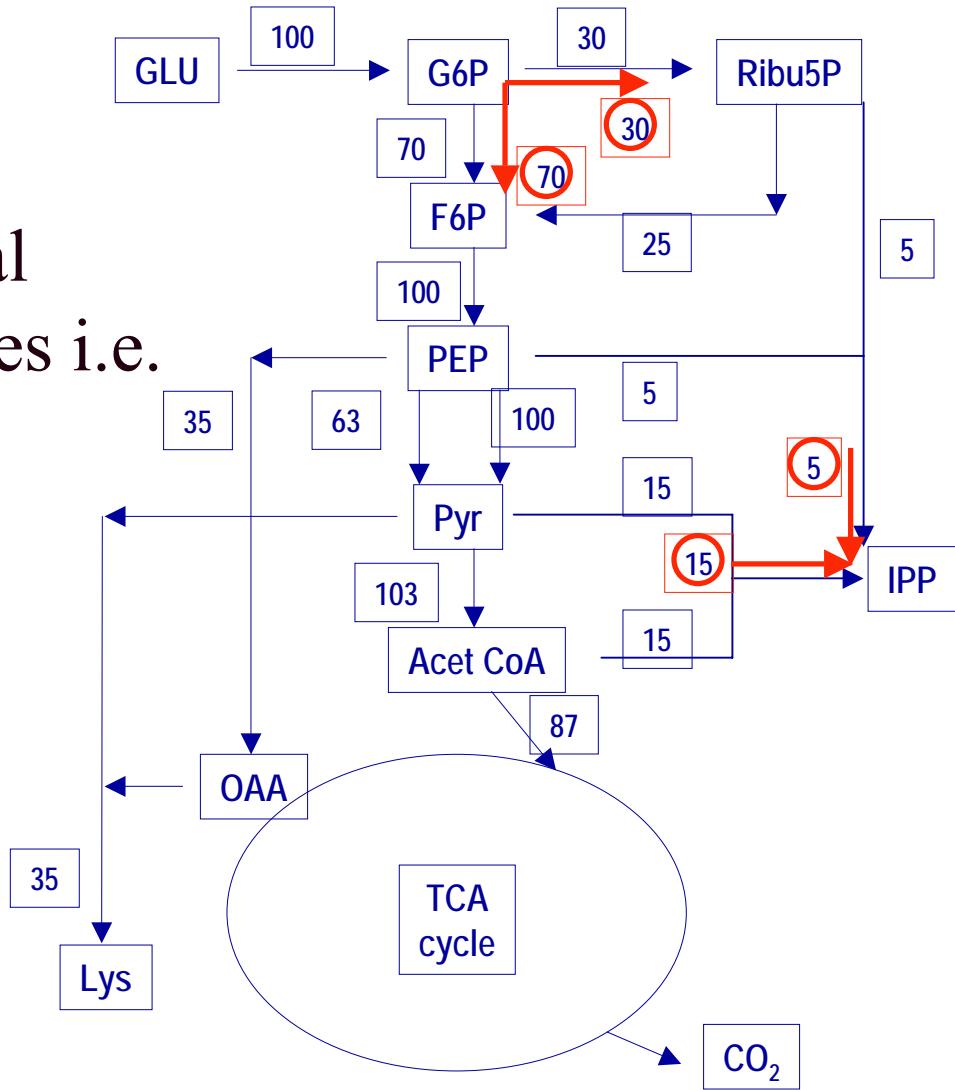
# AS $\alpha$ line – increase in Lochnericine

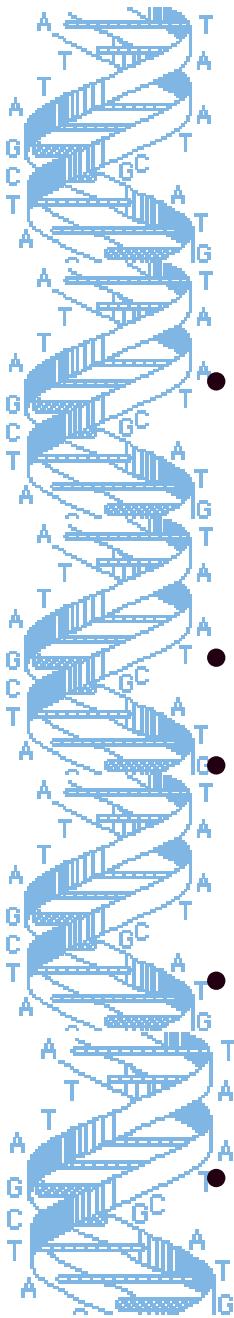




# Metabolic Flux Analysis (MFA)

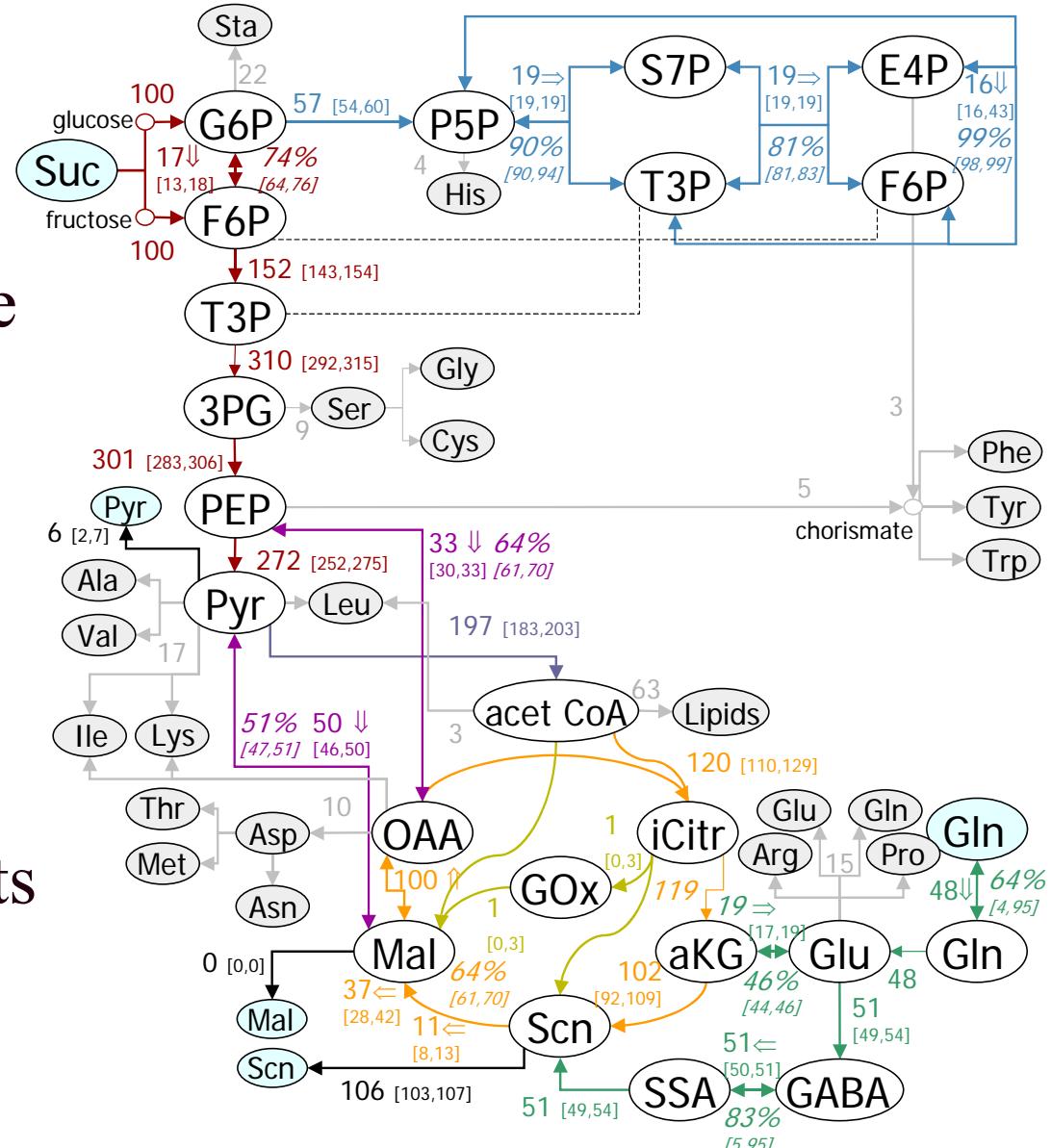
- NMR/GC data
  - provides additional experimental values i.e. flux ratios
  - value shown in microbial studies

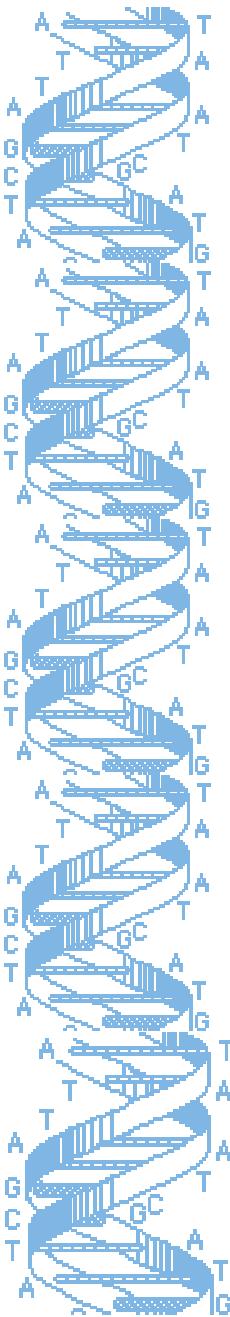




# Flux map

- Network Structure
  - In one or more compartments
- Flux values
- Uncertainty
- Few measurements needed
- Fast computation

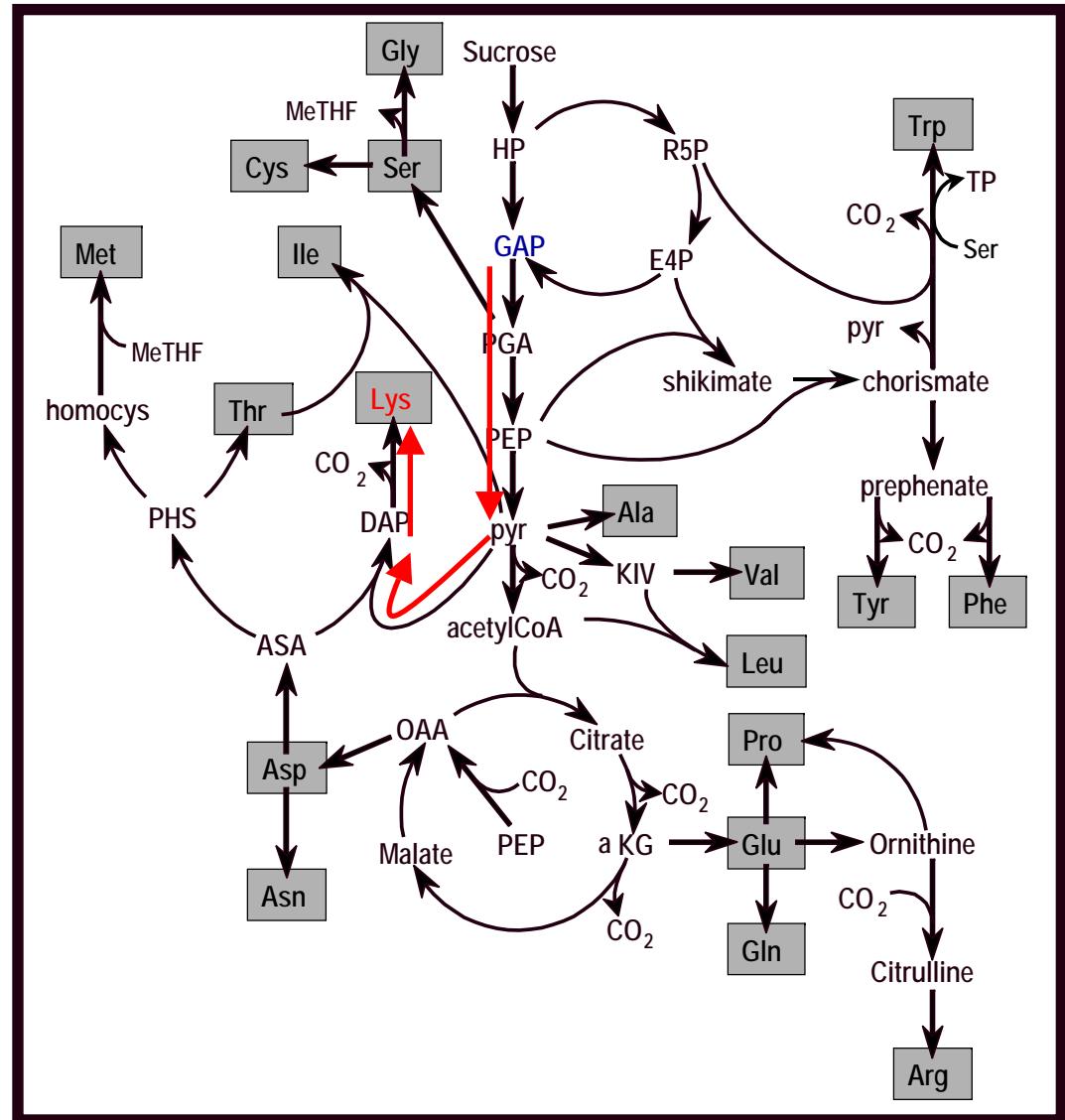


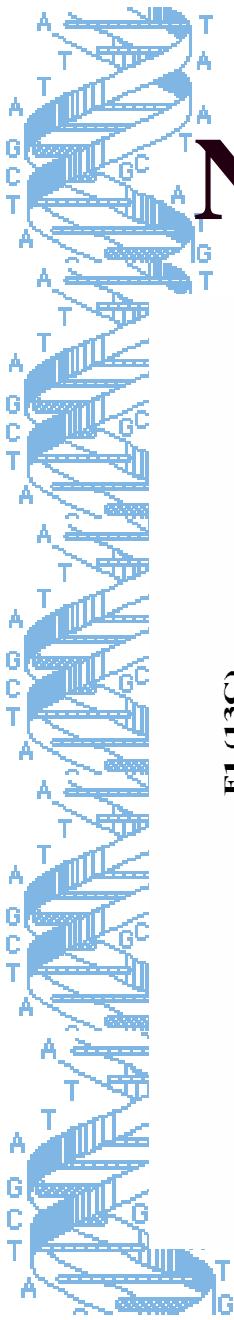


# Bond-labeling experiment (BLE): rationale

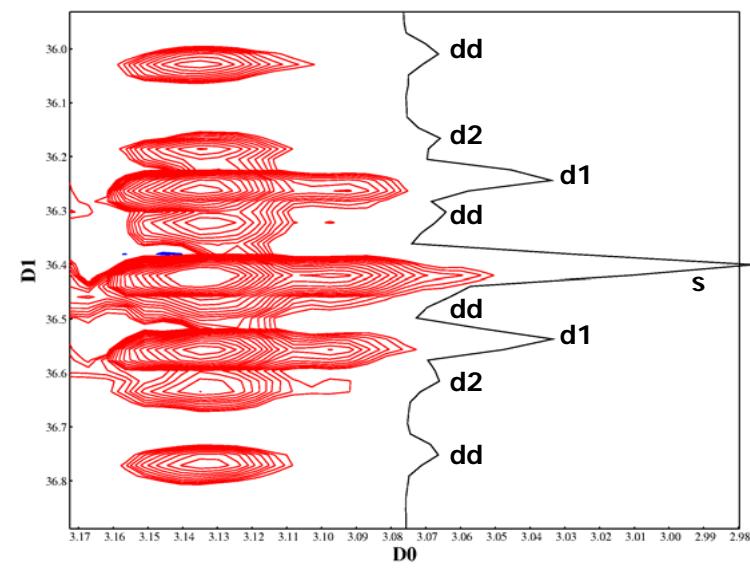
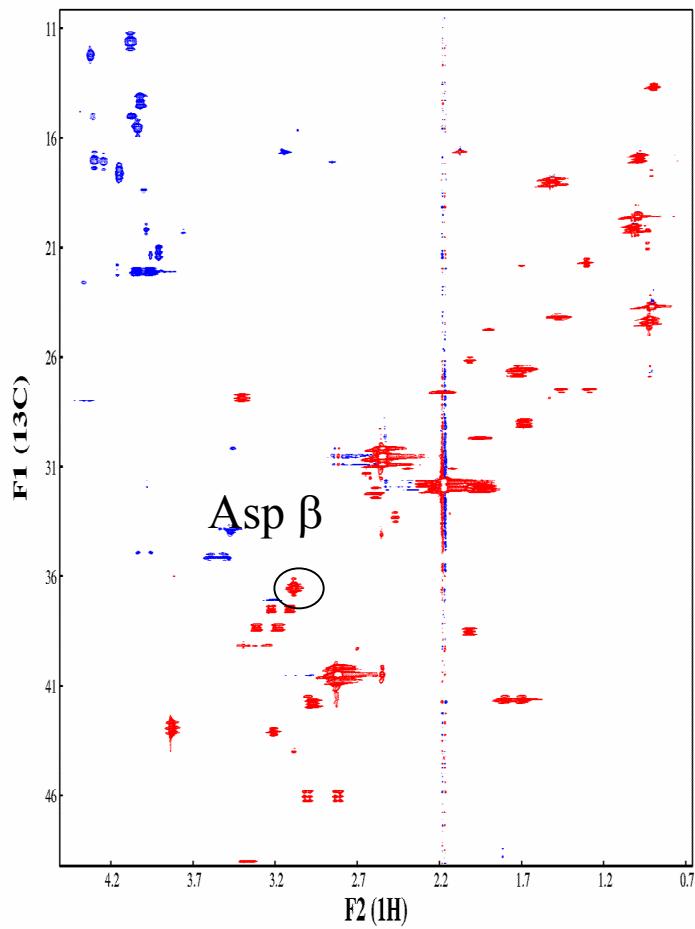
- Amino acids (AAs) reflect the structure of their precursors
- For instance, a part of the **Lys molecule** reflects the structure of **GAP**
- Thus if the **ensemble** of all AAs from an organism is analyzed, it will provide information about the structure of a number of central carbon metabolism precursors (which depends on metabolic flux)

How do we do this?

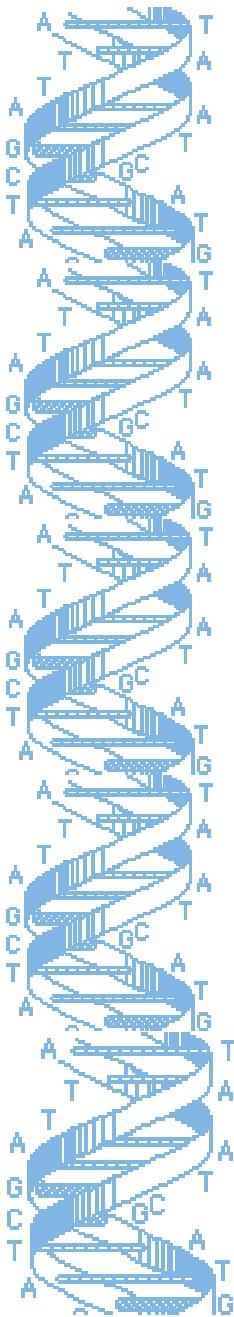




# NMR spectrum of amino acids

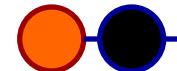


Asp  $\beta$



# $^{13}\text{C}$ fine structures

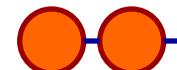
$^{13}\text{C}$  attached to  $^{12}\text{C}$



singlet

*Most likely formed by a biosynthetic bond between a  $^{13}\text{C}$  and a  $^{12}\text{C}$  molecule*

$^{13}\text{C}$  attached to  $^{13}\text{C}$

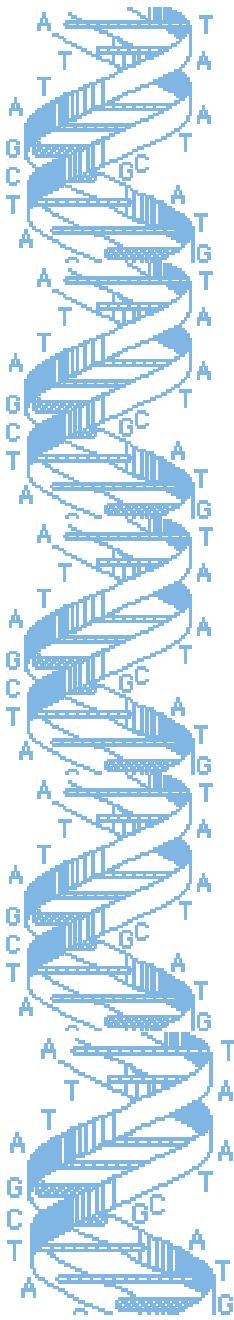


doublet

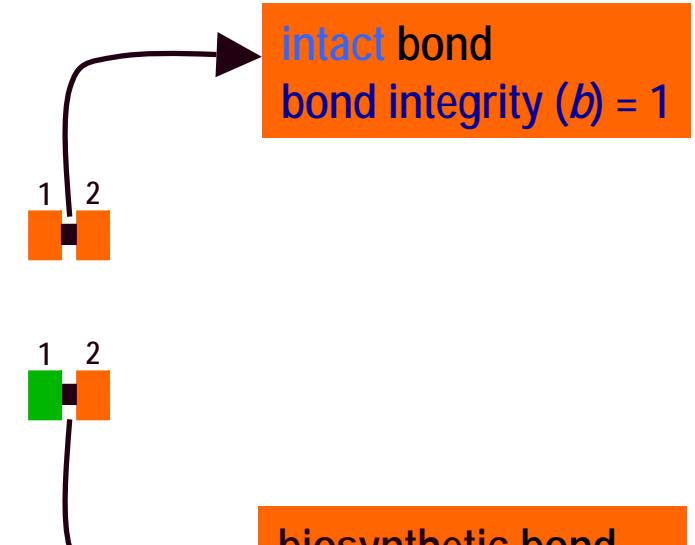
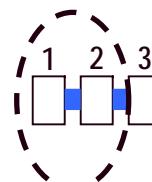
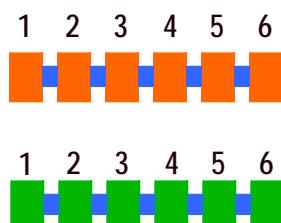
*Most likely formed from an intact  $^{13}\text{C}$  molecule*

Different metabolic histories

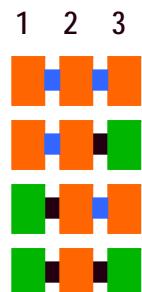
Relative abundance of doublets and singlets represents the relative concentrations of intact and biosynthetic bonds in the same metabolite molecule (e.g. different pathways)



# Bond integrity and bondomers



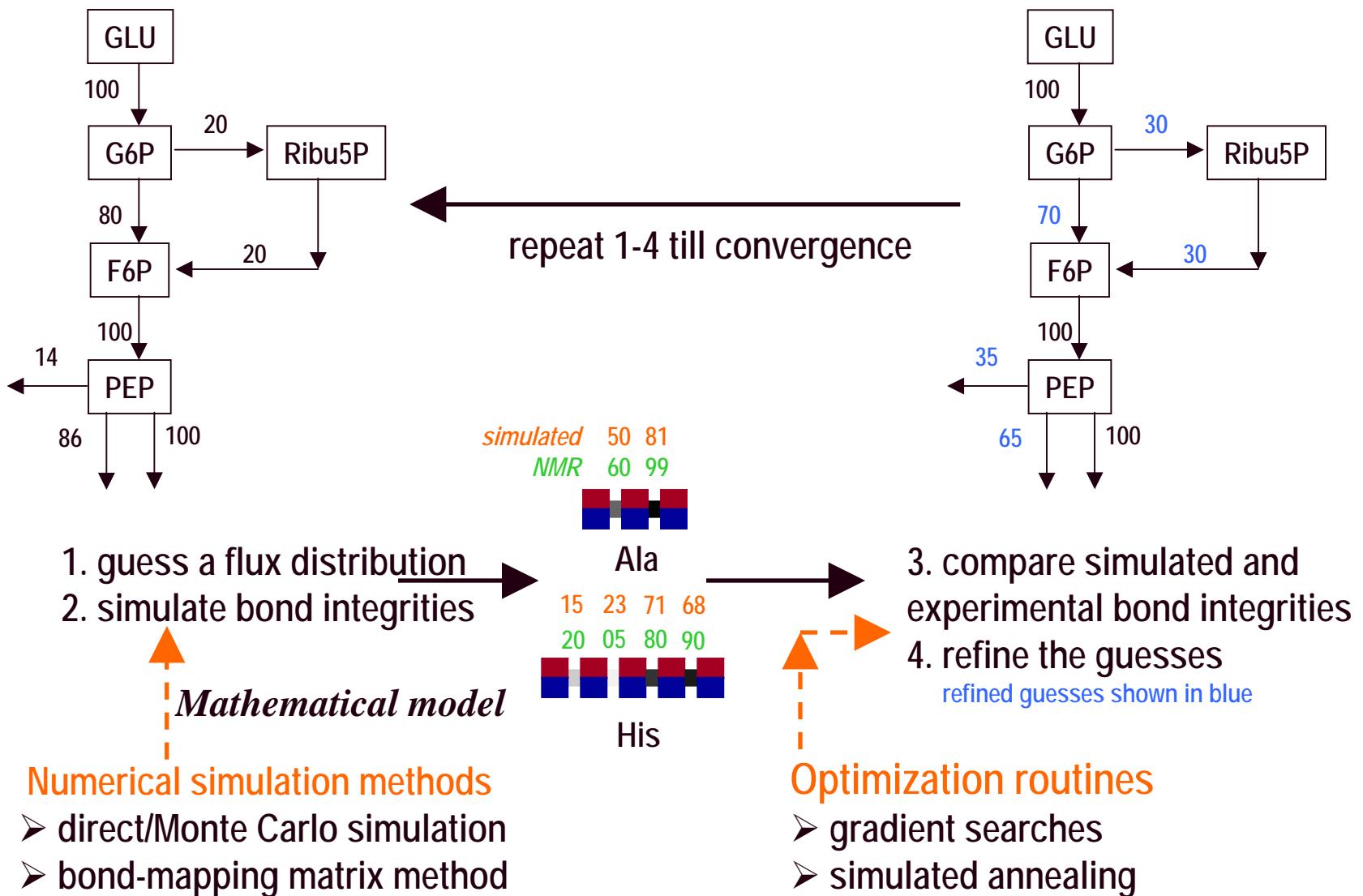
Bondmers of GAP

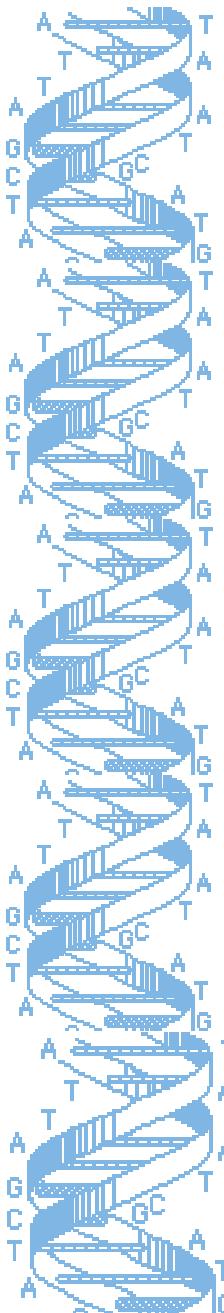


metabolite  
e.g. GAP

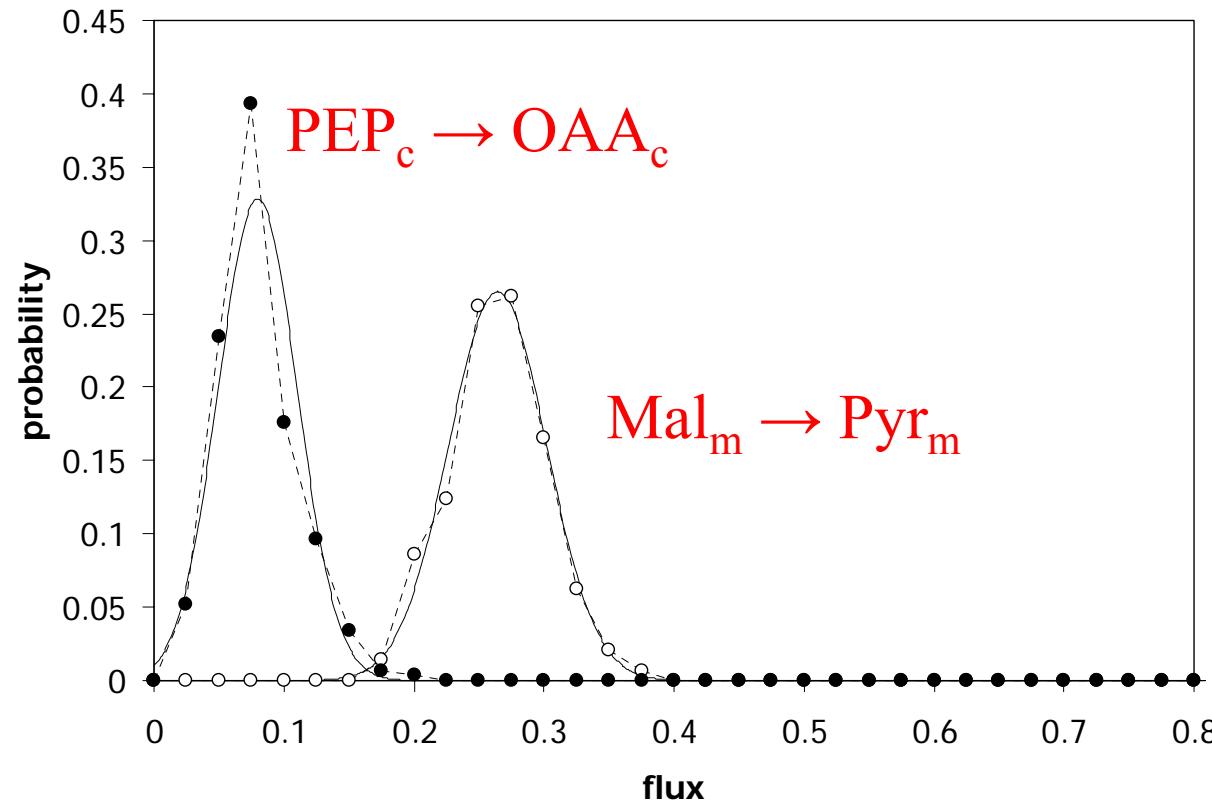
1-1, $\text{GAP}_{4'}$	$4 = 1 \times 2^1 + 1 \times 2^0 (+1)$
1-0, $\text{GAP}_{3'}$	$3 = 1 \times 2^1 + 0 \times 2^0 (+1)$
0-1, $\text{GAP}_{2'}$	$2 = 0 \times 2^1 + 1 \times 2^0 (+1)$
0-0, $\text{GAP}_{1'}$	$1 = 0 \times 2^1 + 0 \times 2^0 (+1)$

# Solution strategy

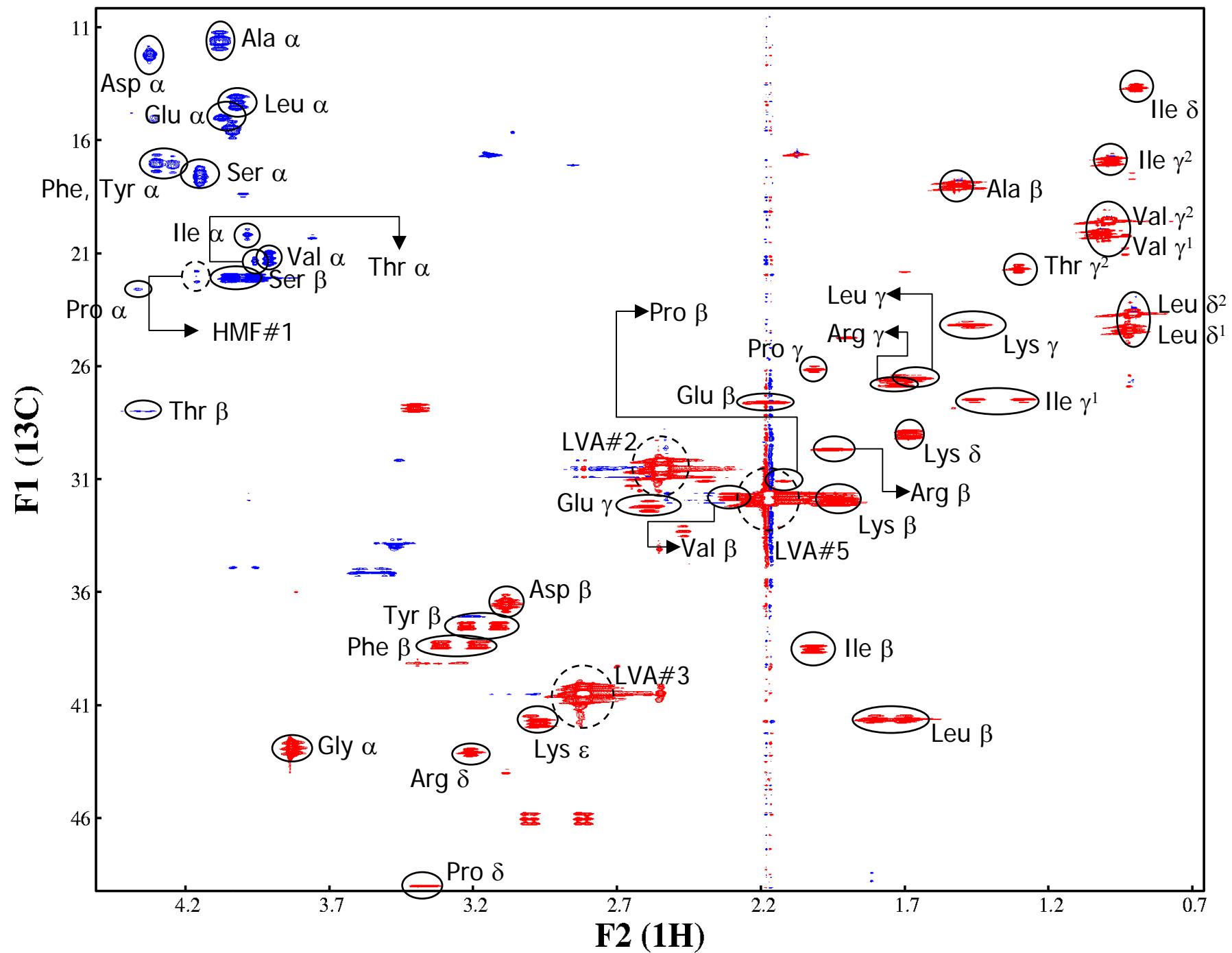




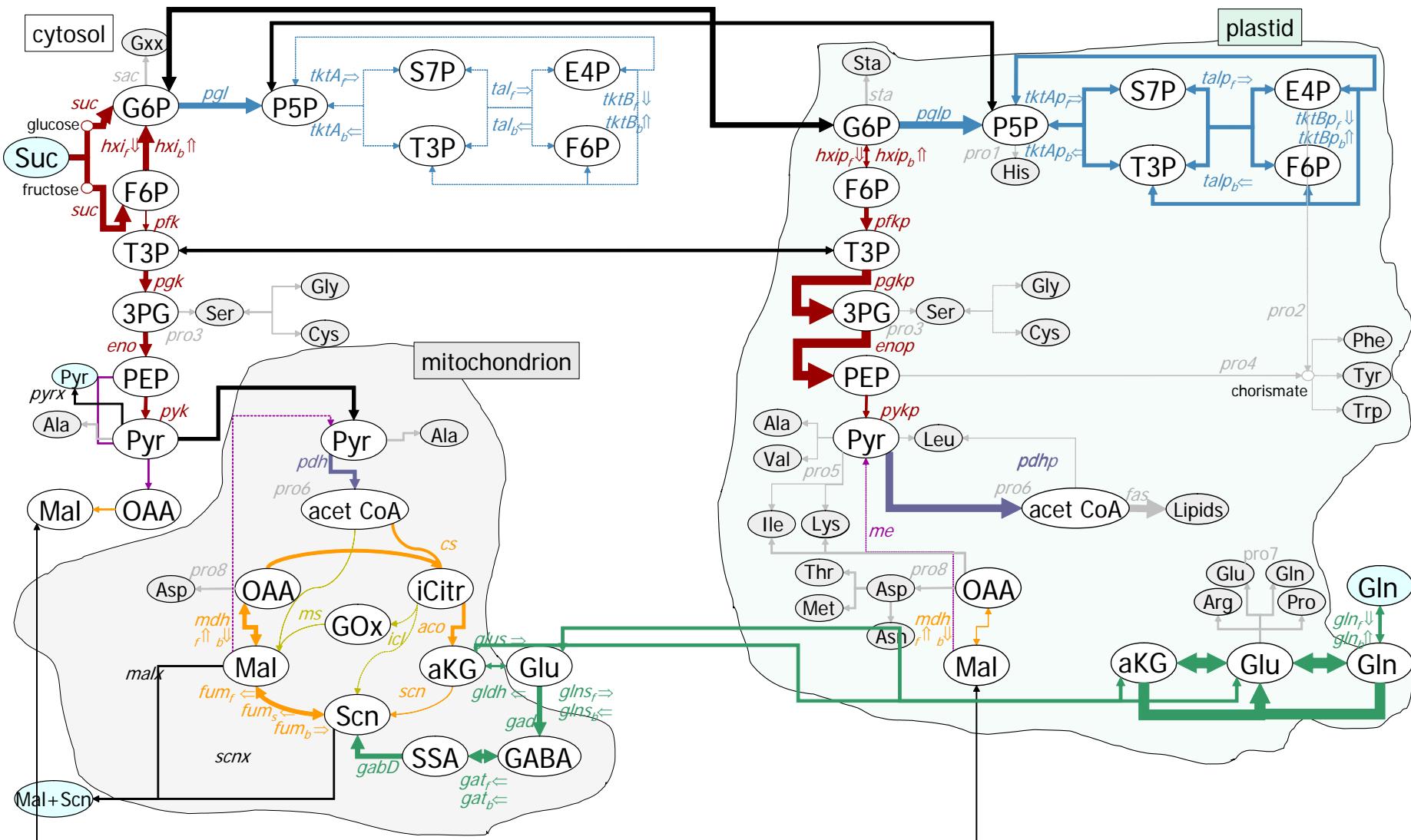
# Statistical analysis of evaluated fluxes



Takes into account experimental error of NMR measurements



# Flux Map of Soybean Embryo Metabolism

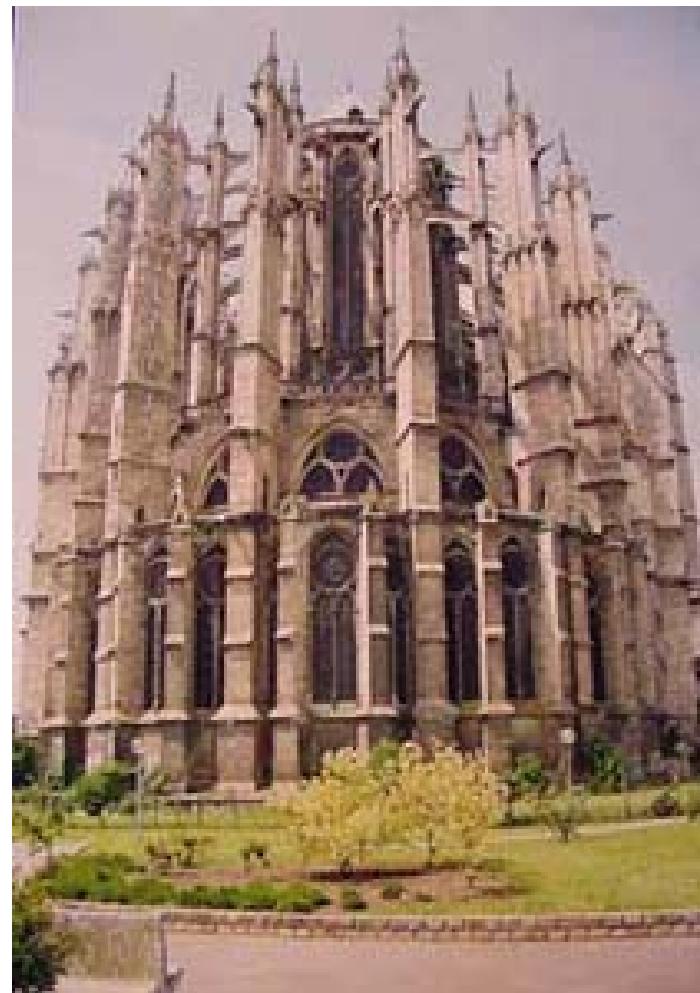


# Impact

- Enhanced tryptophan → essential amino acid in crops for animal consumption
- Systems viewpoint to enhance the overproduction of medicinal metabolites in plants
- Quantitative NMR flux maps in plants are an important tool to be integrated with other systems approaches in metabolic engineering



# Beauvais Cathedral





# Acknowledgments

## Flux Map

- Ganesh Sriram (ChE)
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- Dr. Louisa Tabatabai (Protein Facility)
- Dr. Bruce Fulton (NMR Facility)

## Alkaloids

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- Christie Peebles
- John Morgan
- Sushil Rijhwani
- Sundeep Vani
- Rajiv Bhadra

**Funding:** NSF, Plant Sciences Institute, Cargill



# Panel Session



# *Improving* Plants

- In silico Plant?
- Systems biology
- “Predictive” Metabolic Engineering
  - Iterative cycle of hypothesis testing
- Can we learn some basic design principles by integrating information in subsets in metabolism?



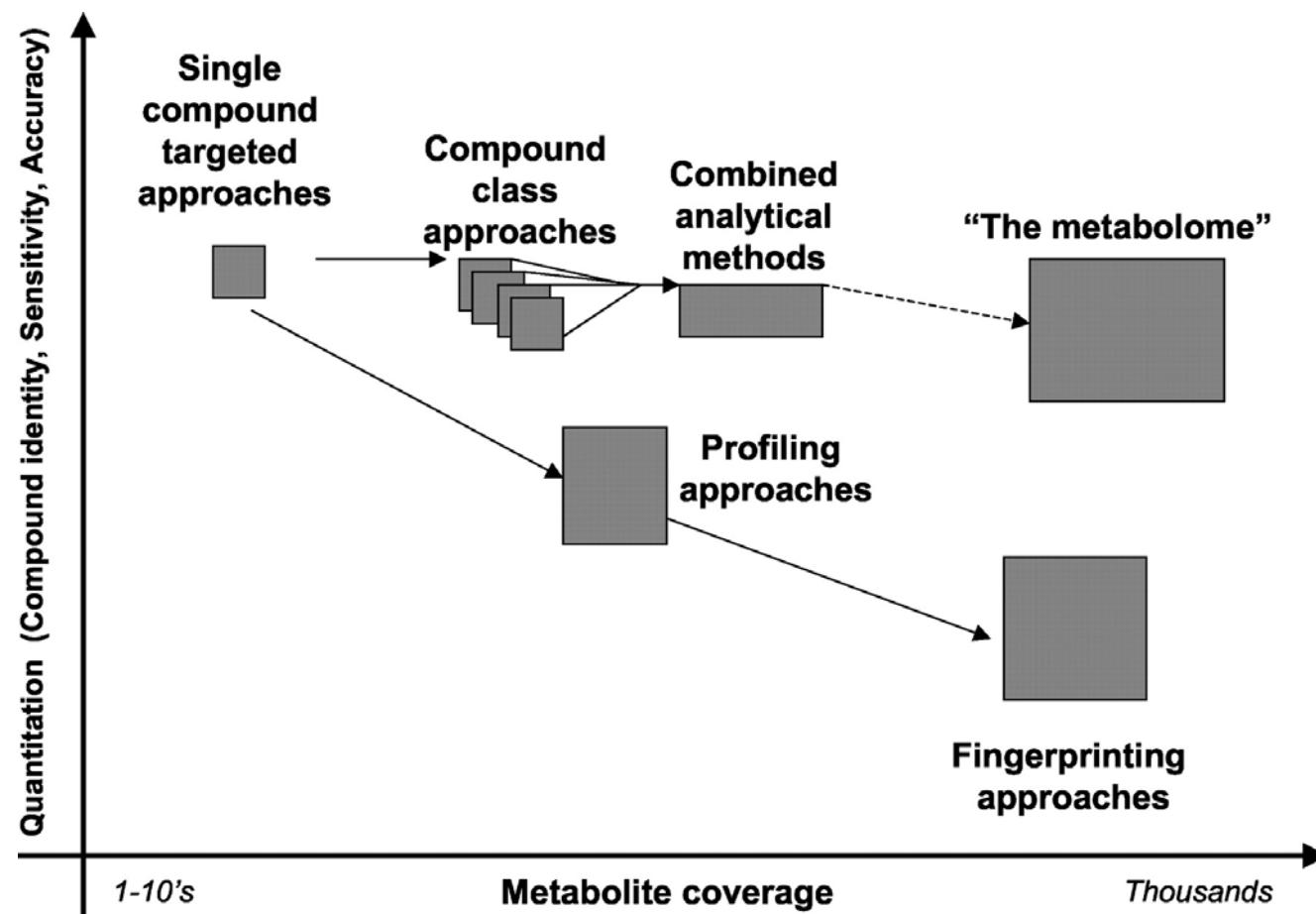
# Challenges – Analysis Side

- **High-throughput techniques**
    - more quantitative
    - proteomics
    - metabolite profiling
    - metabolic flux profiling
  - **Accurate databases with common language**
  - **Theoretical frameworks and tools**
    - Modeling
    - Computational
    - Statistical
    - Visualization





# Approaches to Metabolite Measurements



Sweetlove, Last and Fernie (2003) "Predictive Metabolic Engineering:  
A Goal for Systems Biology" *Plant Phys.* 132: 420-425.